

EFFECT OF CYTOPLASMIC POLYHEDROSIS ON SILKWORM (*BOMBYX MORI* L.) AND ITS TRANSMISSION FROM GENERATION TO GENERATION

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ABSTRACT

The LC_{50} for larval mortality during fifth instar was 0.096×10^6 cytoplasmic polyhedral inclusion bodies ml^{-1} in Pure Mysore and 0.218×10^6 PIBs ml^{-1} in NB_4D_2 silkworms. Besides mortality of silkworms, there was a significant reduction in the cocoon weight, shell weight and shell ratio in the surviving silkworms. The F_1 progeny obtained by selfing the moths emerged from the silkworms inoculated with BmCPV exhibited inferiority in fecundity, hatchability, larval weight, cocoon yield, cocoon weight, shell weight, shell ratio, filament length, denier and effective rate of rearing, when compared to control batches.

Key words : *Bombyx mori*, BmCPV, commercial characters, polyhedrosis.

INTRODUCTION

The silkworm cocoon crops are highly unpredictable due to several factors including various diseases. Of various pathogens, the cytoplasmic polyhedrosis virus (BmCPV) of *Bombyx mori* L. is one of the most widely distributed viruses in the sericultural belt of Karnataka state and causes considerable damage to silk production. In Karnataka alone the infection of BmCPV accounted 27.76% (Samson *et al.*, 1988). In contrast to pebrine (*Nosema bombycis* Nageli), there is no clear-cut evidence about the mode of transmission of BmCPV from generation to generation, or about its effects on the commercial characters of the progeny. However, Hukuhara (1962) and Aruga and Nagashima (1962) reported that the BmCPV is transmitted from generation to generation in *Bombyx mori*. In contrast, surface contamination of eggs in

Pectinophora gossypiella (Bullock *et al.*, 1969), *Heliothis virescens* (Sikorowski *et al.*, 1973 Merry and Dulmage, 1975) is an important means of transmitting the CPV from generation to generation. Neilson (1965) in four species of lepidoptera, Vail *et al.*, (1969) and, Vail and Gouch (1970) in *Trichoplusia ni*, Bullock *et al.*, (1970) in *Pectinophora gossypiella*, Magnoler (1974) in *Parthetria dispar*, Simmons and Sikorowski (1973) and Sikorowski and Thomson (1979) in *Heliothis virescens*, observed the larval mortality, increased larval and pupal duration, reduced larval and pupal weight, reduced moth emergence with deformed, undersized adults, reduced fecundity in the diseased individuals. But, studies on the effects of BmCPV on commercial characters of silkworm *Bombyx mori* and its transmission from generation to generation are rather scarce. Therefore, present

investigation was carried out.

MATERIALS AND METHODS

Two mulberry silkworm races namely Pure Mysore (multivoltine), and NB₄D₂ (bivoltine) at the age of fifth instar first day and a stock of cytoplasmic polyhedrosis virus (BmCPV) were used for the study. The cytoplasmic polyhedral inclusion bodies (PIBs), obtained from Sericulture Department, University of Agricultural Sciences, G.K.V.K., Bangalore, India, were *per orally* inoculated into the silkworm larvae for multiplication. Isolation and purification of PIBs were carried out by following the method described by Balakrishnappa and Honnaiah (1992).

The silkworm rearing was conducted in the laboratory following the method described by Krishnaswami (1978). For determination of breed susceptibility, a total of seven batches, each batch containing 50 worms in triplicate, were inoculated by oral injection with 40 µl of different concentrations of PIBs viz., 6.25 x 10⁶, 3.125 x 10⁶, 1.562 x 10⁶, 0.781 x 10⁶, 0.391 x 10⁶, 0.195 x 10⁶ and 0.0976 x 10⁶ ml⁻¹ polyhedral bodies in 0.75% NaCl solution. The control worms received the same amount of 0.75% NaCl solution only. Later, the worms were allowed to complete larval stage, spinning, pupation and moth emergence. Number of cocoons harvested from each batch were considered for calculation of the breed susceptibility. The moths emerged from BmCPV inoculated larvae were selfed and the F₁ progeny raised were used to study the effects of BmCPV on the economic characters of silkworm. Two doses of BmCPV, viz., 1.562 x 10⁶ ml⁻¹ (T₁) and 3.125 x 10⁶ ml⁻¹ (T₂) were selected for inoculation after preliminary studies on breed susceptibility. The larvae were allowed to complete larval duration, spinning, pupation and moth emergence. Cocoons were harvested, preserved racewise and

treatmentwise at room temperature of 25±2°C and relative humidity of 75±5%. The procedure followed for the preparation of layings & incubation, silkworm rearing and assessment of economic traits are as described by Narasimhanna (1988), Krishnaswami (1978) and Mahesha (1997), respectively. The data derived from the above, experiments were statistically analysed by one way ANOVA (Fisher and Yates, 1953) and Duncan Multiple Range Test (Duncan, 1955). The LC₅₀ values were calculated for BmCPV infection by using probit analysis (Fenney, 1971).

RESULTS AND DISCUSSION

The LC₅₀ for larval mortality during fifth instar was 0.096 ± 0.0024 x 10⁶ PIBs ml⁻¹ in Pure Mysore followed by NB₄D₂ (0.28 ± 0.0027 x 10⁶ PIBs ml⁻¹). This indicates that Pure Mysore was more sensitive to BmCPV when compared with NB₄D₂ race. In Pure Mysore as well as NB₄D₂ races, the viability rate was higher in the control batches and gradual reduction was noticed as the dose of BmCPV increased. According to Bhaskar (1996), the Pure Mysore is sensitive to BmCPV than NB₄D₂ in early stages of larval development. However, contrary to the general trend, the multivoltine Pure Mysore, was found to be more sensitive to BmCPV as compared to a bivoltine NB₄D₂ race. Such a contradiction might have direct relationship to the body weight as well as larval duration; that is, lighter race with longer larval duration (Pure Mysore) is more susceptible to BmCPV as against the heavier race with lesser larval duration (NB₄D₂). Apart from the mortality of silkworms, there was a significant reduction in the cocoon weight, shell weight and shell ratio in the surviving silkworms of both Pure Mysore as well as NB₄D₂ races (Table 1). This might be due to reduced feeding accompanied by the reduced

Table 1. Effect of cytoplasmic polyhedrosis on larval mortality and cocoon characters

| Dose of Bm CPV | No. of cocoon harvested | Race | | | No. of cocoons harvested | NB ₄ D ₂ | | |
|---|-------------------------|-------------------|------------------|-----------------|--------------------------|--------------------------------|------------------|-----------------|
| | | Pure Mysore | | | | NB ₄ D ₂ | | |
| | | Cocoon weight (g) | Shell weight (g) | Shell ratio (%) | | Cocoon weight (g) | Shell weight (g) | Shell ratio (%) |
| (Control) | 46 | 0.958 | 0.139 | 14.51 | 45 | 1.884 | 0.364 | 19.32 |
| C ₁ (0.098x10 ⁶ ml ⁻¹) | 22 | 0.850 | 0.102 | 12.00 | 26 | 1.804 | 0.301 | 16.68 |
| C ₂ (0.195x10 ⁶ ml ⁻¹) | 18 | 0.840 | 0.087 | 10.36 | 19 | 1.786 | 0.287 | 15.79 |
| C ₃ (0.391x10 ⁶ ml ⁻¹) | 13 | 0.821 | 0.081 | 9.87 | 15 | 1.754 | 0.265 | 15.11 |
| C ₄ (0.781x10 ⁶ ml ⁻¹) | 12 | 0.778 | 0.074 | 9.51 | 14 | 1.702 | 0.256 | 15.04 |
| C ₅ (0.562x10 ⁶ ml ⁻¹) | 10 | 0.750 | 0.070 | 9.33 | 12 | 1.671 | 0.251 | 14.96 |
| C ₆ (3.125x10 ⁶ ml ⁻¹) | 7 | 0.733 | 0.069 | 9.41 | 9 | 1.639 | 0.243 | 14.82 |
| C ₇ (6.25x10 ⁶ ml ⁻¹) | 5 | 0.701 | 0.062 | 9.41 | 6 | 1.570 | 0.226 | 14.39 |
| F Ratio | 177.461 | 133.480 | 12.055 | 5.437 | 12.384 | 212.728 | 400.288 | 10.121 |
| Probability | 0.000 | 0.000 | 0.000 | 0.002 | 0.000 | 0.000 | 0.000 | 0.000 |

digestibility and absorption, since the pathogen destroys the midgut tissue. Moreover, the food material absorbed by the pathogen might be utilized for its reproduction and metabolism. In addition, the energy derived from the digested food must have distributed between the host as well as virus for their mutual defense purpose.

The F₁ progeny obtained by selfing the moths emerged from the silkworms inoculated with BmCPV, exhibited inferiority as compared to control batches in almost all the economic characters analysed. The size of the eggs in the silkworm batches treated with BmCPV was small in Pure Mysore, but NB₄D₂ did not show significant reduction in the egg size. Almost all commercial characters like fecundity, hatchability, larval weight, cocoon yield, cocoon weight, shell weight and ratio, filament length, denier and

effective rate of rearing showed reduction when compared to the control sets of both the races (Table 2 and 3). Vail *et al.* (1969) and, Vail and Gouch (1970) in *Trichoplusia ni* made similar observations. Bullock *et al.* (1970) in *Pectinophora gossypiella*, Simmons and Sikorowski (1973) and, Sikorowski and Thomson (1979) in *Heliothis virescens*. The reduction in the egg size and fecundity might be due to the deviation of digested food from normal metabolic state to synthesis of viral proteins and/or due to reduced rate of ingestion, digestion and assimilation of food owing to the malfunction of the midgut of the worms infected with BmCPV. The inferiority in the economic traits of progeny might be due to the following reasons. Firstly, as the egg size was found to be smaller in the BmCPV treated batches, the quantity of yolk, which is

Table 2. Effect of cytoplasmic polyhedrosis on commercial characters of F₁ progeny (Race : Pure Mysore)

| Dose of BmCPV | Fecundity | Hatching (%) | Wt. of 10 V th age larva | | Yield per 10000 larvae brushed | | Single cocoon wt. (g) | Single shell wt (g) | Shell ratio | Filament length (m) | Denier | ERR |
|--|-----------|--------------|-------------------------------------|-----|--------------------------------|-------------|-----------------------|---------------------|-------------|---------------------|--------|--------|
| | | | (g) | (h) | Number | Weight (kg) | | | | | | |
| Control (00) | 504 | 98.49 | 21.52 | 672 | 9400 | 7.445 | 0.856 | 0.114 | 13.32 | 381 | 1.94* | 94.00 |
| T ₁ (1.562x10 ⁶ ml ⁻¹) | 439 | 96.11 | 20.04 | 672 | 9324 | 6.748 | 0.784 | 0.101* | 12.88* | 348 | 1.92* | 93.24* |
| T ₂ (3.125X10 ⁶ ml ⁻¹) | 401 | 95.41 | 19.46 | 672 | 9309 | 6.595 | 0.771 | 0.096* | 12.45* | 321 | 1.92* | 93.09* |
| F Ratio | 42.39 | 97.55 | 114.89 | - | 37.783 | 235.93 | 265.73 | 9.15 | 3.10 | 55.67 | 2.40 | 7.17 |
| Probability | 0.000 | 0.000 | 0.000 | - | 0.000 | 0.000 | 0.000 | 0.015 | 0.119 | 0.000 | 0.172 | 0.026 |

Table 3. Effect of cytoplasmic polyhedrosis on commercial characters of F₁ progeny (Race : NB₄D₂)

| Dose of BmCPV | Fecundity | Hatching (%) | Wt. of 10 V th age larva | | Yield per 10000 larvae brushed | | Single cocoon wt. (g) | Single shell wt (g) | Shell ratio | Filament length (m) | Denier | ERR |
|--|-----------|--------------|-------------------------------------|-----|--------------------------------|-------------|-----------------------|---------------------|-------------|---------------------|--------|---------|
| | | | (g) | (h) | Number | Weight (kg) | | | | | | |
| Control (00) | 580 | 98.11 | 44.05 | 576 | 9418 | 17.800 | 1.950 | 0.376 | 19.28 | 1054 | 2.28 | 94.18 |
| T ₁ (1.562x10 ⁶ ml ⁻¹) | 520* | 95.46 | 41.71 | 576 | 9340 | 16.680 | 1.846 | 0.312 | 16.90* | 924 | 2.21* | 93.40 |
| T ₂ (3.125X10 ⁶ ml ⁻¹) | 496* | 94.01 | 40.09 | 576 | 9311 | 15.955 | 1.774 | 0.284 | 16.00* | 881 | 2.22* | 93.11 |
| F Ratio | 25.880 | 34.215 | 36.238 | - | 162.124 | 431.008 | 403.008 | 31.609 | 12.143 | 16.638 | 21.500 | 271.784 |
| Probability | 0.001 | 0.001 | 0.001 | - | 0.000 | 0.000 | 0.000 | 0.001 | 0.008 | 0.004 | 0.002 | 0.000 |

The variation between control and experimental sets, and among different doses are all significant at 5% level.

* Not significant at 5% level.

reserved for embryonic development, was also reduced; ultimately weak larva may hatch out. Secondly, it is also possible that BmCPV can be transmitted from generation to generation in an occult state, which might render the larva of *Bombyx mori* weak and incapable to perform normal metabolism. Thus, inferior economic characters are produced. The findings of Hukuhara (1961) and Aruga and Nagashima (1962) support such reasoning. They reported that the BmCPV is

transmitted from generation to generation in *Bombyx mori*. In contrast, Sikorowski *et al.* (1973) reported that CPV is transmitted in the tobacco budworm, *Heliothis virescens*, on the surface of the eggs rather than inside them. Bullock *et al.* (1969) also concluded that the surface contamination of eggs is an important means of transmitting the CPV of the pink bollworm, *Pectinophora gossypiella*. However, Sikorowski *et al.* (1971) observed that the virus particles are

present in the haemolymph of *Heliothis virescens* larvae and in the adult. This result suggested that the developing eggs might be under the influence of haemolymph, which contains virus particles. Thus, inferior commercial characters might be due to weak larvae hatched out from the undersized eggs produced under the influence of BmCPV.

Hence, the present investigation clearly indicated that the progeny obtained by selfing the moths emerged from BmCPV inoculated silkworms, exhibited inferiority in almost all characters analysed, and this knowledge can be used in the sericulture industry during the selection of parent seed cocoons for the preparation of disease free layings either for commercial or academic purposes, which have a vital role in the improvement of the industry.

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REFERENCES

- Aruga H. and Nagashima, E. 1962. Generation to generation transmission of the cytoplasmic polyhedrosis virus of *Bombyx mori* (Linneus). J. Insect Pathol., 4 : 313-320.
- Balakrishnappa, Y.K. and Honnaiah, S. 1992. Isolation and purification of cytoplasmic polyhedrosis virus from *Bombyx mori* L.J. Mysore University, Sect. B 32 : 331-333.
- Bhaskar, R.N. 1996. Epizootiological aspects of cytoplasmic polyhedrosis of silkworm *Bombyx mori* L.Ph.D. Thesis, University of Agricultural Sciences, Bangalore, India.
- Bullock, H.R., Mangum, C.L. and Guerra, A.A. 1969. Treatments of eggs of pink ball worm, *Pectinophora gossypiella*, with formaldehyde to prevent infection with a cytoplasmic polyhedrosis virus. J. Invertebrate Pathol., 14 : 271-273.
- Bullock, H.R., Martinier, E. and Stuermer, C.W. 1970. Cytoplasmic polyhedrosis virus and the development and fecundity of pink bollworm. J. Invertebrate Pathol., 15:109-112.
- Duncan, D.B. 1955. Multiple range and Multiple F tests. Biometrics, 11:1-42.
- Fenney, D.J. 1971. Probit analysis. III Edn., 90-233.
- Fisher, A.R. and Yates, F. 1953. Statistical tables for biological, agricultural and medical research. 6th Edn. Longman Group Ltd., England.
- Hukuhara, T. 1962. Generation to generation transmission of the cytoplasmic polyhedrosis virus of silkworm *Bombyx mori* (Linneus). J. Insect Pathol., 4:132-135.
- Krishnaswami, S. 1978. New technology of silkworm rearing. C.S.R. & T.I. Bulletin no. 2, Central Silk Board, India, 1-23.
- Magnoler, A. 1974. Effects of a cytoplasmic polyhedrosis on larval and post larval stages of the Gypsy moth, *Parthetria dispar*. J. Invertebrate Pathol., 23:263-274.
- Mahesha, H.B. 1997. Cytogenetic and biochemical

- studies in a few races of *Bombyx mori* L., treated with cytoplasmic polyhedrosis virus and a mutagen, Ph. D Thesis, University of Mysore, Mysore, India. :41-45.
- Mery, C. and Dulmage, H.T. 1975. Transmission, diagnosis and control of cytoplasmic polyhedrosis virus in colonies of *Heliothis virescens*. J. Invertebrate Pathol., 26:75-79.
- Narasimhanna, M.N. 1988. Manual on silkworm egg production. Central Silk Board, India.
- Neilson, M.M. 1965. Effects of a cytoplasmic polyhedrosis on adult lepidoptera. J. Invertebrate Pathol., 7:306-314.
- Samson, M.V. Baig M., Balavenkatasubbaiah, M., Sharma, S. D., Sasidharan, T.O. and Jolly, M.S. 1988. Infectivity titer of three viral inoculum of cytoplasmic polyhedrosis to silkworm, *Bombyx mori* L. Indian J.Seric., 27 :113-116.
- Sikorowski, P.P., Andrews, G.L. and Broome J.R. 1973. Trans-ovum transmission of a cytoplasmic polyhedrosis virus of *Heliothis virescens* (Lepidoptera : Noctuidae). J. Invertebrate Pathol., 21
- Sikorowski, P.P., Andrews G.L and Broome J.R. 1971. Presence of cytoplasmic polyhedrosis virus in the haemolymph of *Heliothis virescens* larvae and adults. J.Invertebrate pathol., 18:167-168.
- Sikorowski, P.P. and Thomson, A.C. 1979. Effects of cytoplasmic polyhedrosis virus on diapausing *Heliothis virescens*. J.Invertebrate Pathol., 33: 66-70.
- Simmons, C.L. and Sikorowski, P.P. 1973. A laboratory study of the effects of cytoplasmic polyhedrosis virus on *Heliothis virescens* (Lepidoptera : Noctuidae). J. Invertebrate Pathol., 22: 237-244.
- Vail, P.V., Hall, I.M. and Gouch, D. 1969. Influence of a cytoplasmic polyhedrosis on various developmental stages of the cabbage looper. J. Invertebrate Pathol., 14 : 271-272.
- Vail, P.A. and Gouch, D. 1970. Effects of cytoplasmic polyhedrosis on adult cabbage loopers and their progeny. J. Invertebrate Pathol., 15 : 397-400.